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Patentanmeldung Nr. Patent application No. Demande de brevet n°

03027514.3

Der Präsident des Europäischen Patentamts;
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For the President of the European Patent Office

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PCT/EP2004 / 013535

Anmeldung Nr:
Application no.: 03027514.3
Demande no:

Anmeldetag:
Date of filing: 28.11.03
Date de dépôt:

Anmelder/Applicant(s)/Demandeur(s):

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Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:
(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.
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Method for preventing and treating diabetes using DG119

In Anspruch genommene Priorität(en) / Priority(ies) claimed / Priorité(s)
revendiquée(s)
Staat/Tag/Aktenzeichen/State/Date/File no./Pays/Date/Numéro de dépôt:

Internationale Patentklassifikation/International Patent Classification/
Classification internationale des brevets:

A61K38/00

Am Anmeldetag benannte Vertragstaaten/Contracting states designated at date of
filing/Etats contractants désignées lors du dépôt:

AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HU IE IT LU MC NL
PT RO SE SI SK TR LI

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Patentanwälte
European Patent Attorneys · European Trademark Attorneys

EPO - Munich
47

28. Nov. 2003

Unser Zeichen:

32043P EP/WWld

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28. Nov. 2003

Method for preventing and treating diabetes using DG119

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Method for preventing and treating diabetes using DG119**Description**

The present invention relates generally to methods for preventing and/or treating pancreatic disorders, particularly those related to diabetes, by administering a DG119-1 product or an agonist thereof and/or an antagonist to a DG119-2 product.

BACKGROUND OF THE INVENTION

The pancreas is an exocrine gland that secretes digestive enzymes directly into the digestive tract as well as an endocrine gland that secretes hormones into the blood stream. The exocrine function is assured by acinar and centroacinar cells that secrete various digestive enzymes via intercalated ducts into the duodenum. The functional unit of the endocrine pancreas is the islet of Langerhans. Islets are scattered throughout the exocrine portion of the pancreas and are composed of four main cell types: alpha-, beta-, delta- and PP-cells (reviewed for example in Kim & Hebok, 2001, Genes Dev. 15: 111-127). Beta-cells produce insulin, represent the majority of the endocrine cells and form the core of the islets, while alpha-cells secrete glucagon and are located in the periphery. Delta-cells and PP-cells are less numerous and secrete somatostatin and pancreatic polypeptide, respectively. Recently, cells producing the neuropeptide Ghrelin have been found in pancreatic islets (Wierup et al., 2002, Regul Pept. 107:63-9.).

Early pancreatic development has been well studied in different species, including chicken, zebrafish, and mice (for a detailed review, see Kim & Hebok, 2001, supra). The pancreas develops from distinct dorsal and ventral anlagen. Pancreas development requires specification of the pancreas anlage along both anterior-posterior and dorsal-ventral axes. A number of transcription factors, that are critical for proper pancreatic development have been identified (see Kim & Hebok, 2001, supra; Wilson et al., 2003, Mech

Dev. 120: 65-80).

In humans, the acinar and ductal cells retain a significant proliferative capacity that can ensure cell renewal and growth, whereas the islet cells become
5 mostly mitotically inactive. This is in contrast to rodents where beta-cell replication is an important mechanism in the generation of new beta cells. It has been suggested, that during embryonic development, pancreatic islets of Langerhans originate from differentiating duct cells or other cells with epithelial morphology (Bonner-Weir & Sharma, 2002, J Pathol. 197: 519-526;
10 Gu et al., 2003, Mech Dev. 120: 35-43). In adult humans, new beta-cells arise in the vicinity of ducts (Butler et al., 2003, Diabetes 52: 102-110; Bouwens & Pipeleers 1998, Diabetologia 41: 629-633). However, also an intra-islet location or an origin in the bone marrow has been suggested for precursor cells of adult beta-cells (Zulewski et al., 2001, Diabetes 50: 521-533;
15 lanus et al., 2003, J Clin Invest. 111: 843-850). Pancreatic islet growth is dynamic and responds to changes in insulin demand, for example during pregnancy or due to changing body weight during childhood. In adults, there is a good correlation between body mass and islet mass (Yoon et al., 2003, J Clin Endocrinol Metab. 88: 2300-2308).

20 Blood vessels play an important role in organ development (Matsumoto et al., 2001, Science 294: 559-563; Lammert et al., 2003, Mech Dev. 120(1): 59-64). For the formation of pancreatic buds from primitive endoderm, the presence of adjacent blood vessels is necessary (Lammert et al., 2001,
25 Science 294: 564-567). The authors also show that blood vessels can induce the expression of insulin in primitive endoderm in tissue conjugation experiments. Furthermore, mice genetically engineered to develop additional blood vessels in their pancreas show a greatly increased islet number. There is a close association between duct-like structures and blood
30 vessels in fetal mouse pancreas, suggesting that blood vessels may play an important role in beta cell neogenesis from ducts (reviewed in Cleaver & Melton, 2003, Nat Med. 9(6):661-668). Thus, it is of great importance to identify signals produced by endothelial cells which can regulate or induce

the generation of functional cell types during embryogenesis and/or adult regeneration. Of particular interest are signals which control the formation of new pancreatic beta cells due to their potential relevance for diabetes therapies.

5

Pancreatic beta-cells secrete insulin in response to rising glucose levels and other secretagogues such as arginine. Insulin amongst other hormones plays a key role in the regulation of the fuel metabolism. Insulin leads to the storage of glycogen and triglycerides and to the synthesis of proteins. The entry of
10 glucose into muscles and adipose cells is stimulated by insulin. In patients who suffer from diabetes mellitus type I or LADA (latent autoimmune diabetes in adults (Pozzilli & Di Mario, 2001, Diabetes Care. 8:1460-67) beta-cells are destroyed due to autoimmune attack. The amount of insulin produced by the remaining pancreatic islet cells is too low, resulting in elevated blood glucose
15 levels (hyperglycemia). In diabetes type II, liver and muscle cells lose their ability to respond to normal blood insulin levels (insulin resistance). High blood glucose levels (and also high blood lipid levels) in turn contribute to an impairment of beta-cell function and to an increase in beta-cell apoptosis. It is interesting to note that the rate of beta-cell neogenesis does not appear to
20 change significantly in type II diabetics (Butler et al., 2003 supra), thus causing a reduction in total beta-cell mass over time. Eventually the application of exogenous insulin becomes necessary in type II diabetics.

Improving metabolic parameters such as blood sugar and blood lipid levels
25 (e.g. through dietary changes, exercise, medication or combinations thereof) before beta-cell mass has fallen below a critical threshold leads to a relatively rapid restoration of beta-cell function. However, even after such a treatment the pancreatic endocrine function would remain impaired due to the only slightly increased regeneration rate. Treatments which increase the
30 rate of neogenesis will have a beneficial effect due to enhanced insulin secretory capacity.

In type I diabetics, where beta-cells are being destroyed by autoimmune

attack, treatments have been devised which modulate the immune system and may be able to stop or strongly reduce islet destruction (Raz et al., 2001, Lancet 358: 1749-1753; Chatenoud et al., 2003, Nat Rev Immunol. 3: 123-132; Homann et al., Immunity. 2002, 3:403-15). However, due to the relatively slow
5 regeneration of human beta-cells such treatments can only be successful if they are combined with agents that can stimulate beta-cell regeneration.

A variety of model organisms has been used to study the formation of beta cells and to analyze the effect of treatments aimed at the improvement of
10 diabetic conditions. Zebrafish has become a popular model vertebrate for the study of developmental processes as well as for pharmacological and toxicological studies over the last decade (Rubinstein, 2003, Curr Opin Drug Discov Devel. 6(2):218-23; Grunwald & Eisen, 2002, Nat Rev Genet. 3(9): 717-24). In this organism, large numbers of transparent embryos which
15 rapidly develop outside of their mother are readily available. Transgenic lines expressing fluorescent proteins under the control of tissue-specific promoters allow to rapidly assess the effects of pharmacological treatments or gene loss- and gain-of-function treatments. Zebrafish islets contain the same celltypes in a similar spatial organization as mammalian islets. A large
20 number of genes which control pancreatic development in mammals also control pancreatic development in zebrafish (Biemar et al., 2001, Dev Biol. 230(2): 189-203; Ober et al., 2003, Mech Dev. 120(1): 5-18). Suppressing gene function in zebrafish embryos using antisense oligonucleotides, modified Peptide Nucleic Acids (mPNAs) or other antisense compounds with
25 good efficiency and specificity yields phenotypes which are usually indistinguishable from genetic mutants in the same gene (Nasevicius et al., Nat Genet. 2000 26(2):216-20; Effimov et al., NAR 26; 566-575; Urtishak et al., 5th international conference on zebrafish development and genetics, Madison/WI 2002, abstr. #17). Thus, zebrafish embryos represent a relevant
30 model to identify genes or compounds which control beta cell formation in humans.

Diabetes is a very disabling disease, because today's common anti-diabetic

drugs do not control blood sugar levels well enough to completely prevent the occurrence of high and low blood sugar levels. Chronically elevated blood sugar levels are toxic and cause long-term complications such as renopathy, retinopathy, neuropathy and peripheral vascular disease. There is also a host of related conditions, such as obesity, hypertension, heart disease and hyperlipidemia, for which persons with diabetes are substantially at risk.

Apart from the impaired quality of life for the patients, the treatment of diabetes and its long term complications presents an enormous financial burden to our healthcare systems with rising tendency. Thus, for the prevention or treatment of diabetes mellitus type I, LADA, and diabetes mellitus type II there is a strong need in the art to identify factors that induce regeneration of pancreatic insulin producing beta-cells.

SUMMARY OF THE INVENTION

In this invention, we disclose a novel and so far unknown use for DG119 proteins as factors secreted from blood vessels within the pancreas to regulate endocrine development. The DG119 proteins regulate or induce the generation of functional cell types during embryogenesis and/or adult regeneration. DG119 proteins control the formation of new pancreatic beta cells.

In particular, DG119-1 is disclosed as positive regulator of insulin-producing cells. Thus, DG119-1 stimulates the formation or regeneration of insulin producing cells, particularly beta-cells. Thus, DG 119-1 and certain modulators (e.g. agonists, stimulators) can be used in the treatment and/or prevention of diseases caused by, accompanied by and/or associated with dysfunctions of pancreatic cells, particularly pancreatic beta-cells, such as diabetes mellitus type I, LADA, and diabetes mellitus type II.

DG119-2 is disclosed herein as negative regulator of insulin-producing cells. Thus, certain modulators (e.g., antagonists, inhibitors) of DG119-2 stimulate

the formation or regeneration of insulin producing beta-cells and can be used in the treatment and/or prevention of diseases caused by, accompanied by and/or associated with dysfunctions of pancreatic cells, particularly pancreatic beta-cells, such as diabetes mellitus type I, LADA, and diabetes mellitus type II.

More particularly, the present invention provides methods for treating patients suffering from a disease linked to functionally impaired and/or reduced numbers of pancreatic islet cells, particularly insulin producing beta-cells, by administering a therapeutically effective amount of a DG119-1 product or an agonist/stimulator thereof and/or an antagonist/inhibitor of a DG119-2 product. Functional impairment or loss of pancreatic islet cells may be due to autoimmune attack in diabetes type I or LADA, or due to cell degeneration in progressed diabetes type II. The methods of the present invention may also be used to treat patients at risk to develop degeneration of insulin producing beta-cells to prevent the start or progress of such a process.

Numerous additional aspects and advantages of the invention will become apparent to those skilled in the art upon consideration of the following description of the Figures and detailed description of the invention which describes presently preferred embodiments thereof.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows internal organs of 17 day old fish embryos.

The fish embryos carry a reporter construct consisting of a fluorescent protein cDNA functionally linked to insulin promoter sequences. Green fluorescence in these fish marks regions of endogenous zebrafish insulin expression. At 17 days of development, a zebrafish pancreas includes one bigger islet and several smaller islets (see arrowheads). Also shown are the gall bladder and the gut of the 17 day old fish embryos.

Figure 2 shows that administration of DG119-1 increases the main islet (Brockmann body) size of 17 day old fish.

Main islets of five 17 day old fish embryos injected either with mRNA of mouse DG119-1 (200 µg/µl) or with control mRNA at the same concentration are shown. Note the relative increase of the mean Brockmann body size in DG119-1 injected embryos.

Figure 3 shows that administration of DG119-1 increases the size of small islets in 17 day old fish.

Isolated gastrointestinal tracts of five 17 day old fish embryos injected at one-cell-stage either with mRNA of mouse DG119-1 or with control mRNA at the same concentration (200 µg/µl) are shown. Brockmann bodies and smaller islets are in green. Note the relative increase of 'small islets' size in DG119-1 injected embryos.

Figure 4 shows that DG119-1 and DG119-2 are expressed in pancreatic blood vessels in mammals.

Figure 4A shows a cryosection of a pancreas from a 17,5 day embryonic mouse that was analysed by in hybridization with antisense mouse DG119-1 probe (dark blue) and stained with anti-insulin antibodies (red) and nuclear dye DAPI (light blue). DG119-1 is mainly located to pancreatic blood vessels (note anuclear erythrocytes within the vessel).

Figure 4B shows a cryosection of a pancreas from 17,5 day embryonic mouse which was analysed by in hybridization with antisense mouse DG119-2 probe (dark). DG119-2 is located to pancreatic blood vessels, similar to DG119-1.

Figure 5 shows that loss of DG119-2 function increases islet size and insulin expression in zebrafish.

One cell stage zebrafish carrying the transgene with insulin regulatory sequences linked to a fluorescent protein cDNA were either injected with antisense oligonucleotides to DG119-2A or with control oligonucleotides.

Pictures of the islet were taken at 24 hpf (A,B). Note the relative

increase of islet size in DG119-2A antisense injected fish (**FIGURE 5B**) compared with the control (**FIGURE 5A**).

Figure 5C shows the relative expression level of Insulin, PDX1 and Pax4 RNA in 48 Hpf. DG119-2A (light grey columns) or control (black and white columns) antisense-injected embryos measured using quantitative RT-PCR procedure. Note three fold increase in insulin RNA content in DG119-2A antisense injected fish. **Figure 5C** shows that these enlarged islets generate higher insulin levels.

Figure 6. Sequences of zebrafish DG119 proteins.

Figure 6a. Sequence of zebrafish DG119-1a protein (719 amino acids, shown in the one-letter-code)(SEQ ID NO:1)

Figure 6b. Sequence of zebrafish DG119-1b protein (594 amino acids, shown in the one-letter-code) (SEQ ID NO:2)

Figure 6c. Sequence of zebrafish DG119-2a protein (146 amino acids, shown in the one-letter-code; C-terminus incomplete) (SEQ ID NO:3)

Figure 6d. Sequence of zebrafish DG119-2b protein (287 amino acids, shown in the one-letter-code; C-terminus incomplete) (SEQ ID NO:4)

FIGURE 7. Sequence alignment of DG119 proteins from human, mouse, and zebrafish.

DESCRIPTION OF THE INVENTION

Before the present invention is described, it is understood that all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs.

In the present invention the term "beta-cell regeneration" refers to the restoration of normal beta-cell function by increasing the number of functional insulin secreting beta-cells and/or by restoring normal function in functionally impaired beta-cells.

As used herein, the term "DG119 product" includes proteins such as purified

natural, synthetic, or recombinant DG119-1 or DG119-2 and variants thereof. Variants include insertion, substitution and deletion variants and chemically modified derivatives. Variants also include recombinant proteins, for example but not limited to hybrids or fusions of DG119-1 or DG119-2 and other proteins. Also included are proteins or peptides substantially homologous to the human DG119-1 or DG119-2 protein having the amino acid sequence published as GenBank Accession Number XP_034000 (DG119-1) or GenBank Accession Number NP_872293 (DG119-2). The term "DG119 product" also includes nucleic acids, e.g. RNA or DNA coding for the above described DG119-1 or DG119-2 protein product. The term "DG119 product" also includes DG119-1 or DG119-2 homodimers or heterodimers of a DG119-1 or DG119-2 protein product and another protein.

The term "substantially homologous" as used herein means having a degree of homology to the biologically active DG119 protein product having the amino acid sequence published as GenBank Accession Number XP_034000 (DG119-1) or GenBank Accession Number NP_872293 (DG119-2), that is preferably in excess of 70%, most preferably in excess of 80%, and even more preferably in excess of 90% or 95%. The degree of homology between the mouse and the human protein is about 91 %, and it is contemplated that preferred mammalian DG119 proteins will have a similarly high degree of homology. Also included are proteins which are hybrids between DG119-1 and another protein which retain the stimulatory effect on islet cell formation found in DG119-1. The percentage of homology or percent identity between a DG119 product and a human DG119 protein or nucleic acid may be determined according to standard procedures, e.g. by using the BLAST algorithm. Preferably, it is calculated as the percentage of nucleotide or amino acid residues found in the smaller of the two sequences that align with identical nucleotide or amino acid residues in the sequence being compared, when four gaps in a length of 100 nucleotides or amino acids may be introduced to assist in that alignment. Also included as substantially homologous is any DG119 protein product which may be isolated by virtue of cross-reactivity with antibodies to the DG119 protein

product or whose genes may be isolated through hybridization with the gene or with segments of the gene encoding the DG119 protein product.

As used herein, the term "agonist/stimulator" of a DG119-1 product refers to any substance that is inducing or stimulating the expression and/or function of DG119-1.

As used herein, the term "antagonists to DG119-2" refers to any substance that is interfering with the expression and/or function of DG119-2. It includes any effectors or modulators of DG119-2, e.g. antagonists or inhibitors. In particular, the "antagonists to DG119-2" can be fragments or otherwise modified parts of DG119-2. Fragments for example could one of two modules of the protein; e.g. the cystein-rich domain (amino acids 30-118 in human DG119-2) or the C-terminal olfactomedin domain (amino acids 447-704 in human DG119-2) or peptides derived therefrom could be used as antagonist to DG119-2. In addition to fragmentation, antagonists to DG119-2 can also be obtained by changing single amino acids or by chemical modifications of the protein. The "antagonists to DG119-2" can include any effectors, e.g. receptors, enzymes, proteins, ligands, agents, or substrates that either directly or indirectly modulate (inhibit) the action of DG119-2 protein products. The "antagonists to DG119-2" can include effectors/modulators of DG119-2 polynucleotides and/or polypeptides, antibodies, biologically active nucleic acids, such as antisense molecules, RNAi molecules or ribozymes, aptamers, peptides or low-molecular weight organic compounds recognizing said DG119-2 polynucleotides or polypeptides.

The term "biologically active" as used herein means that the DG119-1 protein product or an agonist thereof stimulates and/or induces the differentiation of insulin producing cells and/or promotes the protection, survival, or regeneration of islet cells.

In connection with the present invention, the term 'progenitor cells' relate to undifferentiated cells capable of being differentiated into insulin producing

cells. The term particularly includes stem cells, i.e. undifferentiated or immature embryonic, adult, or somatic cells that can give rise to various specialized cell types. The term "stem cells" can include embryonic stem cells (ES) and primordial germ cells (EG) cells of human or animal origin.

5 Isolation and culture of such cells is well known to those skilled in the art (see, for example, Thomson et al., 1998, Science 282:1145-1147; Shambloott et al., 1998, Proc. Natl. Acad. Sci. USA 95:13726-13731; US 6,090,622; US 5,914,268; WO 0027995; Notarianni et al., 1990, J. Reprod. Fert. 41:51-56; Vassilieva et al., 2000, Exp. Cell. Res. 258:361-373). Adult or somatic stem
10 cells have been identified in numerous different tissues such as intestine, muscle, bone marrow, liver, and brain. WO 03/023018 describes a novel method for isolating, culturing, and differentiating intestinal stem cells for therapeutic use. In the pancreas, several indications suggest that stem cells are also present within the adult tissue (Gu & Sarvetnick, 1993, Development 118:33-46; Bouwens, 1998, Microsc Res Tech 43:332-336;
15 Bonner-Weir, 2000, J. Mol. Endocr. 24:297-302).

Embryonic stem cells can be isolated from the inner cell mass of pre-implantation embryos (ES cells) or from the primordial germ cells found in
20 the genital ridges of post-implanted embryos (EG cells). When grown in special culture conditions such as spinner culture or hanging drops, both ES and EG cells aggregate to form embryoid bodies (EB). EBs are composed of various cell types similar to those present during embryogenesis. When cultured in appropriate media, EB can be used to generate in vitro
25 differentiated phenotypes, such as extraembryonic endoderm, hematopoietic cells, neurons, cardiomyocytes, skeletal muscle cells, and vascular cells. We have previously described a method that allows EB to efficiently differentiate into insulin-producing cells (as described in patent application PCT/EP02/04362, published as WO 02/086107 and by Blyszczuk et al.,
30 2003, Proc Natl Acad Sci U S A. 100(3):998-1003, which are incorporated herein by reference).

In this invention, we disclose a novel and so far unknown use for DG119-1 to

stimulate and/or induce the formation or regeneration of insulin producing beta-cells and thus, a use in the treatment and prevention of diseases going along with impaired beta-cell function, for example but not limited to diabetes mellitus.

5

The present invention is based on the finding that DG119-1 stimulates the formation of insulin producing islets. Furthermore, this invention discloses that antagonists to DG119-2 stimulate the formation of islets that are insulin producing. Thus, a therapeutically effective amount of DG119-1 product or
10 an agonist thereof or an antagonist to a DG119-2 product may be administered to promote the regeneration of pancreatic beta-cells or to promote the differentiation or formation of insulin-producing cells from progenitor cells in vitro or in vivo. The present invention further relates to applications in the medical field that directly arise from the method of the
15 invention. Additionally, the present invention relates to applications for the identification and characterization of compounds with therapeutic medical effects or toxicological effects that directly arise from the method of the invention.

20 According to this invention the DG119-1 product or agonist or the antagonist to a DG119-2 product may be administered

- i) as a pharmaceutical composition e.g. enterally, parenterally or topically, preferably directly to the pancreas,
 - ii) via implantation of DG119-1 protein product or antagonist to DG119-2
25 product expressing cells, and/or
 - iii) via gene therapy
- as described in more detail below.

Further, the DG119-1 or DG119-2 expression level or function in a patient
30 might be influenced by a pharmaceutically active substance administered

- i) as a pharmaceutical composition e.g. enterally, parenterally or topically, preferably directly to the pancreas,
- ii) via cell based therapy and/or

iii) via gene therapy
as described in more detail below.

5 The DG119-1 product or the pharmaceutically active substance influencing the DG119-1 or DG119-2 expression level or function may be administered in the above described manner alone or in combination with another pharmaceutical composition useful to treat beta-cell degeneration, for example hormones, growth factors or immune modulating agents.

10 A DG119-1 product or an agonist thereof or an antagonist of a DG119-2 product may be administered in patients suffering from a disease going along with impaired beta-cell function, for example but not limited to diabetes type I, LADA, or progressed diabetes type II. It is further contemplated that the above compounds may be administered preventively to patients at risk to
15 develop beta-cell degeneration, like for example but not limited to patients suffering from diabetes type II or LADA in early stages. A variety of pharmaceutical formulations and different delivery techniques are described in further detail below.

20 The invention further comprises the use of cells with activated pancreatic genes, e.g. as described in WO 03/023018, which is herein incorporated by reference. Examples of preferred pancreatic genes are Pdx1, Pax4, Pax6, neurogenin3 (ngn3), Nkx6.1, Nkx6.2, Nkx2.2, HB9, Beta2/NeuroD, Isl1, HNF1-alpha, HNF-1 beta and HNF3 of human or animal origin. The
25 pancreatic genes may be introduced into the cells by transfection or transduction, e.g. transfection of progenitor or stem cells or transduction of pancreatic duct and islet cells (Noguchi H., et al., 2003, Diabetes 52: 1732-1737).

30 DG119 products, e.g. DG119 protein products, are preferably produced via recombinant techniques because such methods are capable of achieving high amounts of protein at a great purity, but are not limited to protein products expressed in bacterial, plant, mammalian, or insect cell systems.

DG119 Protein Product

Recombinant DG119-1 or DG119-2 protein product forms include glycosylated and non-glycosylated forms of the protein. In general, recombinant techniques involve isolating the genes encoding for DG119-1 or
5 DG119-2 protein product, cloning the gene in suitable vectors and/or cell types, modifying the gene if necessary to encode a desired variant, and expressing the gene in order to produce the DG119-1 or DG119-2 protein product.

10 Alternatively, a nucleotide sequence encoding the desired DG119-1 or DG119-2 protein product may be chemically synthesized. It is contemplated that a DG119-1 or DG119-2 protein product may be expressed using nucleotide sequences that vary in codon usage due to the degeneration of the genetic code or allelic variations or alterations made to facilitate
15 production of the protein product by the selected cell.

The DG119-1 or DG119-2 protein products according to this invention may be isolated or generated by a variety of means. Exemplary methods for producing DG119 protein products, vectors, host cells, and culture growth
20 conditions for the expression of DG119-1 or DG119-2 protein, as well as methods to synthesize variants of DG119-1 or DG119-2 protein product are known to those skilled in the art.

DG119-1 or DG119-2 protein product variants are prepared by introducing
25 appropriate nucleotide changes into the DNA encoding the polypeptide or by in vitro chemical synthesis of the desired polypeptide. It will be appreciated by those skilled in the art that many combinations of deletions, insertions, and substitutions can be made resulting in a protein product variant presenting DG119-1 or DG119-2 biological activity. Mutagenesis
30 techniques for the replacement, insertion or deletion of one or more selected amino acid residues are well known to one skilled in the art. DG119-1 or DG119-2 substitution variants have at least one amino acid residue of the human or mouse DG119-1 or DG119-2 amino acid sequence removed and

a different residue inserted in its place. Such substitution variants include allelic variants, which are characterized by naturally occurring nucleotide sequence changes in the species population that may or may not result in an amino acid change.

5 Chemically modified derivatives of DG119-1 or DG119-2 protein products also may be prepared by one of skill in the art given the disclosures herein. The chemical moieties most suitable for derivatization include water soluble polymers. A water soluble polymer is desirable because the protein to which
10 it is attached does not precipitate in an aqueous environment, such as a physiological environment. Preferably, the polymer will be pharmaceutically acceptable for the preparation of a therapeutic product or composition. One skilled in the art will be able to select the desired polymer based on such considerations as whether the polymer/protein conjugate will be used
15 therapeutically, and if so, the desired dosage, circulation time, resistance to proteolysis, and other considerations. A particularly preferred water-soluble polymer for use herein is polyethylene glycol. Attachment at residues important for receptor binding should be avoided if receptor binding is desired. One may specifically desire an N-terminal chemically modified
20 protein.

The present invention contemplates use of derivatives which are prokaryote-expressed DG119-1 or DG119-2, or variants thereof, linked to at least one polyethylene glycol molecule, as well as use of DG119-1 or DG119-2, or
25 variants thereof, attached to one or more polyethylene glycol molecules via an acyl or alkyl linkage.

The present invention also discloses use of derivatives which are prokaryote-expressed DG119-1 or DG119-2, or variants thereof, linked to at
30 least one hydrophobic residue, for example fatty acid molecule, as well as use of DG119-1 or DG119-2, or variants thereof, attached to one or more hydrophobic residues. For example, patent application published as WO 03/010185, which is hereby incorporated by reference, describes a method

for producing acylated polypeptides in transformed host cells by expressing a precursor molecule of the desired polypeptide which are then to be acylated in a subsequent in vitro step.

5 **Polynucleotides Encoding DG119 Protein Product**

The present invention further provides polynucleotides that encode DG119-1 or DG119-2 protein products whether recombinantly produced or naturally occurring.

10 A nucleic acid sequence encoding a DG119-1 or DG119-2 protein product, can readily be obtained in a variety of ways, including, without limitation, chemical synthesis, cDNA or genomic library screening, expression library screening, and/or PCR amplification of cDNA. These methods and others useful for isolating such nucleic acid sequences are set forth, for example,
15 by Sambrook et al. (Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989), by Ausubel et al., eds (Current Protocols in Molecular Biology, Current Protocols Press, 1994), and by Berger and 2 5 Kimmel (Methods in Enzymology: Guide to Molecular Cloning Techniques, vol. 152, Academic Press, Inc., San Diego, CA, 1987).
20 Chemical synthesis of a nucleic acid sequence which encodes a DG119 protein product can also be accomplished using methods well known in the art, such as those set forth by Engels et al. (Angew. Chem. Intl. Ed., 28:716-734, 3 0 1989).

25 Included within the scope of this invention are DG119-1 or DG119-2 product polynucleotides with the native signal sequence and other pre-pro sequences as well as polynucleotides wherein the native signal sequence is deleted and replaced with a heterologous signal sequence. The heterologous signal sequence selected should be one that is recognized and
30 processed, i.e., cleaved by a signal peptidase, by the host cell. For prokaryotic host cells that do not recognize and process the native DG119-1 or DG119-2 signal sequence, the signal sequence may be substituted by a prokaryotic signal sequence selected, for example, from the group of the

alkaline phosphatase, penicillinase, or heat-stable enterotoxin 11 leaders. For yeast secretion, the native DG119-1 or DG119-2 signal sequence may be substituted by the yeast invertase, alpha factor, or acid phosphatase leaders. In mammalian cell expression the native signal sequence is satisfactory, although other mammalian signal sequences may be suitable. Expression and cloning vectors generally include a nucleic acid sequence that enables the vector to replicate in one or more selected host cells.

DG119 Pharmaceutical Compositions

Pharmaceutical compositions comprising a DG119-1 product or an agonist thereof or an antagonist to a DG119-2 product typically include a therapeutically effective amount of the active ingredient in admixture with one or more pharmaceutically and physiologically acceptable formulation. In addition to the active ingredients, pharmaceutical compositions may contain suitable pharmaceutically acceptable carriers comprising excipients and auxiliaries, which facilitate processing of the active compounds into preparations, which can be used pharmaceutically. Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing Co., Easton, Pa.), the disclosure of which is hereby incorporated by reference.

Once the therapeutic composition has been formulated, it may be stored in sterile vials as a solution, suspension, gel, emulsion, solid, or dehydrated or lyophilized powder. Such formulations may be stored either in a ready to use form or in a form, e.g., lyophilized, requiring reconstitution prior to administration. The optimal pharmaceutical formulations will be determined by one skilled in the art depending upon considerations such as the route of administration and desired dosage. Such formulations may influence the physical state, stability, rate of in vivo release, and rate of in vivo clearance of the present DG119-1 or DG119-2 proteins, variants and derivatives. Other effective administration forms, such as slow-release formulations, inhalant mists, or orally active formulations are also envisioned. For example, in a sustained release formulation, the active ingredient may be

bound to or incorporated into particulate preparations of polymeric compounds (such as polylactic acid, polyglycolic acid, etc.) or liposomes.

Administration/Delivery

5 The active ingredient may be administered by any suitable means, e.g. enterally or parenterally or topically directly to the pancreas, as known to those skilled in the art. The specific dose may be calculated according to considerations of body weight, body surface area or organ size. Further refinement of the calculations necessary to determine the appropriate
10 dosage for treatment involving each of the above mentioned formulations is routinely made by those of ordinary skill in the art and is within the ambit of tasks routinely performed. Appropriate dosages may be ascertained through use of the established assays for determining dosages utilized in conjunction with appropriate dose-response data. The final dosage regimen involved in a
15 method for treating the above described conditions will be determined by the attending physician, considering various factors which modify the action of drugs, e.g., the age, condition, body weight, sex and diet of the patient, the severity of any infection, time of administration and other clinical factors. As studies are conducted, further information will emerge regarding the
20 appropriate dosage levels for the treatment of various diseases and conditions.

It is envisioned that the continuous administration or sustained delivery of the active ingredient may be advantageous for a given treatment. While
25 continuous administration may be accomplished via a mechanical means, such as with an infusion pump, it is contemplated that other modes of continuous or near continuous administration may be practiced. For example, chemical derivatization or encapsulation may result in sustained release forms of the protein having the effect of continuous presence, in
30 predictable amounts, based on a determined dosage regimen. Thus, the active ingredients include proteins derivatized or otherwise formulated to effectuate such continuous administration.

Cell therapy, e.g. pancreatic implantation of cells producing DG119-1 protein product or antagonists to DG119-2 protein product, is also contemplated. This embodiment would involve implanting cells capable of synthesizing and secreting a biologically active form of DG119-1 protein product or antagonists to DG119-2 protein product into patients. Such DG119-1 protein product or antagonists to DG119-2 protein product-producing cells may be cells that are natural producers of DG119-1 protein product or antagonists to DG119-2 protein product or may be cells that are modified to express such proteins. Modified cells include recombinant cells whose ability to produce a DG119-1 protein product or an antagonist to DG119-2 protein product has been augmented by transformation with a gene encoding the desired DG119-1 protein product or an antagonist to a DG119-2 product in a vector suitable for promoting its expression and secretion. In order to minimize a potential immunological reaction in patients being administered DG119-1 product or an antagonist to a DG119-2 product of a foreign species, it is preferred that the cells be of human origin and produce a human DG119-1 protein product or human antagonist to a DG119-2 protein product. Likewise, it is preferred that recombinant cells are transformed with an expression vector containing a gene encoding a human DG119-1 protein product or with an inhibitory nucleic acid for the DG119-2 product, e.g. an antisense molecule, a ribozyme or an RNAi molecule or a DNA molecule coding therefor. Implanted cells may be encapsulated to avoid infiltration of surrounding tissue. Human or nonhuman animal cells may be implanted in patients in biocompatible, semipermeable polymeric enclosures or membranes that allow release of the active ingredient but that prevent destruction of the cells by the patient's immune system or by other detrimental factors from the surrounding tissue.

Alternatively, cells may be introduced into a patient in need intraportally via a percutaneous transhepatic approach using local anaesthesia. Such surgical techniques are well known in the art and can be applied without any undue experimentation, see Pyzdrowski et al, 1992, New England J. Medicine 327:220-226; Hering et al., Transplantation Proc. 26:570-571,

1993; Shapiro et al., New England J. Medicine 343:230-238, 2000.

Further, the invention relates to a cell preparation comprising differentiated stem cells exhibiting insulin production, e.g. an insulin-producing cell line obtainable by the method described above. The insulin-producing cells may exhibit a stable or a transient expression of at least one gene involved in beta-cell differentiation. The cells are preferably human cells that are derived from human stem cells. For therapeutic applications the generation of autologous human cells from adult stem cells of a patient is especially preferred. However, the insulin producing cells may also be derived from nonautologous cells. If necessary, immune rejection may be avoided by encapsulation, immunosuppression and/or modulation or due to non-immunogenic properties of the cells. The insulin producing cells preferably exhibit characteristics that closely resemble naturally occurring beta-cells, e.g. an enhanced insulin production by a factor of at least 2, preferably at least 3 after addition of 27.7 mM glucose.

The cell preparation of the invention is preferably a pharmaceutical composition comprising the cells together with pharmacologically acceptable carriers, diluents and/or adjuvants. The pharmaceutical composition is preferably used for the prevention or treatment of pancreatic diseases, e.g. diabetes.

According to the present invention, the functional cells treated with a DG119-1 product or an agonist thereof or an antagonist to a DG119-2 product are transplanted preferably intrahepatic, directly into the pancreas of an individual in need, or by other methods. Alternatively, such cells are enclosed into implantable capsules that can be introduced into the body of an individual, at any location, more preferably in the vicinity of the pancreas, or the bladder, or the liver, or under the skin. Methods of introducing cells into individuals are well known to those of skill in the art and include, but are not limited to, injection, intravenous or parenteral administration. Single, multiple, continuous or intermittent administration can be effected. The cells

can be introduced into any of several different sites, including but not limited to the pancreas, the abdominal cavity, the kidney, the liver, the celiac artery, the portal vein or the spleen. The cells may also be deposited in the pancreas of the individual.

5

The methodology for the membrane encapsulation of living cells is familiar to those of ordinary skill in the art, and the preparation of the encapsulated cells and their implantation in patients may be accomplished without undue experimentation. See, e.g. U.S. Patent Numbers 4,892,538, 5,011,472, and
10 5,106,627, each of which is specifically incorporated herein by reference. A system for encapsulating living cells is described in PCT Application WO 91/10425 of Aebischer et al., specifically incorporated herein by reference. See also, PCT Application WO 91/10470 of Aebischer et al., Winn et al., Exper. Neurol., 113:322-329, 1991, Aebischer et al., Exper. Neurol., 11
15 1:269-275, 1991; Tresco et al., ASAIO, 38:17-23, 1992, each of which is specifically incorporated herein by reference. Techniques for formulating a variety of other sustained- or controlled-delivery means, such as liposome carriers, bio-erodible particles or beads and depot injections, are also known to those skilled in the art.

20

In another embodiment gene therapy ex vivo is envisioned, i.e. the patient's own cells may be transformed ex vivo to produce DG119-1 protein product or a protein stimulating DG119-1 expression or a protein or a substance, e.g. an inhibitory nucleic acid, inhibiting DG119-2 expression and would be
25 directly reimplanted. For example, cells retrieved from the patient may be cultured and transformed with an appropriate vector. After an optional propagation/expansion phase, the cells can be transplanted back into the same patient's body, particularly the pancreas, where they would produce and release the desired protein product. Delivery by transfection and by
30 liposome injections may be achieved using methods, which are well known in the art. Any of the therapeutic methods described above may be applied to any suitable subject including, for example, mammals such as dogs, cats, cows, horses, rabbits, monkeys, and most preferably, humans.

DG119-1 product gene therapy in vivo is also envisioned, by introducing the gene coding for DG119-1 protein product into targeted pancreas cells via local injection of a nucleic acid construct or other appropriate delivery
5 vectors. (Hefti, J. Neurobiol., 25:1418-1435, 1994). For example, a nucleic acid sequence encoding a DG119-1 protein product may be contained in an adenoassociated virus vector or adenovirus vector for delivery to the pancreas cells. Alternative viral vectors include, but are not limited to, retrovirus, herpes simplex virus and papilloma virus vectors. Physical
10 transfer, either in vivo or ex vivo as appropriate, may also be achieved by liposome-mediated transfer, direct injection (naked DNA), receptor-mediated transfer (ligand-DNA complex), electroporation, calcium phosphate precipitation or microparticle bombardment (gene gun).

15 Immunosuppressive drugs, such as cyclosporin, can also be administered to the patient in need to reduce the host reaction versus graft. Allografts using the cells obtained by the methods of the present invention are also useful because a single healthy donor could supply enough cells to regenerate at least partial pancreas function in multiple recipients.

20 In one embodiment of the invention, administration of a DG119-1 product and/or an agonist thereof or an antagonist of a DG119-2 product in a pharmaceutical composition to a subject in need thereof, particularly a human patient, leads to an at least partial regeneration of pancreatic cells. Preferably,
25 these cells are insulin producing beta-cells that will contribute to the improvement of a diabetic state. With the administration of this composition e.g. on a short term or regular basis, an increase in beta-cell mass can be achieved. This effect upon the body reverses the condition of diabetes partially or completely. As the subject's blood glucose homeostasis improves, the
30 dosage administered may be reduced in strength. In at least some cases further administration can be discontinued entirely and the subject continues to produce a normal amount of insulin without further treatment. The subject is thereby not only treated but could be cured entirely of a diabetic condition.

However, even moderate improvements in beta-cell mass can lead to a reduced requirement for exogenous insulin, improved glycemic control and a subsequent reduction in diabetic complications. In another example, the compositions of the present invention will also have efficacy for treatment of patients with other pancreatic diseases such as pancreatic cancer, dysplasia, or pancreatitis, if beta-cells are to be regenerated.

In a further embodiment, the present invention allows the production of cells for the identification and/or characterisation of compounds which stimulate beta-cell differentiation, insulin secretion and/or glucose response, more particularly of compounds which increase the DG119-1 and/or decrease the DG119-2 expression level or function. This method is particularly suitable for in vivo testing for diagnostic applications and drug development or screening. The compound of interest is added to suitable cells and DG119-1 and/or DG119-2 expression or function is determined. Alternatively, a compound of interest is added to a DG119-1, DG119-1 agonist or DG119-2 antagonist treated cell and the effect on cell differentiation and/or insulin production is determined. Preferably, differentiated insulin-producing cells are used. Insulin levels in treated cells can be determined, e.g. quantified by Enzyme Linked Immunoabsorbent Assay (ELISA) or Radio Immuno Assay (RIA). Using this method, a large number of compounds can be screened and compounds that increase DG119-1 expression or support the activity of DG119-1 or compounds that decrease DG119-2 expression or inhibit the activity of DG119-2 leading to beta-cell differentiation and/or an increase in insulin secretion can be identified readily.

In a high-throughput screening method, the cells are transfected with a DNA construct, e.g. a viral or non-viral vector containing a reporter gene, e.g. the lacZ gene or the GFP gene, under regulatory control of a promoter of a gene involved in beta-cell differentiation, e.g. preferably a Pax4 promoter. The transfected cells are divided into aliquots and each aliquot is contacted with a test substance, e.g. candidate 1, candidate 2, and candidate 3. The activity of the reporter gene corresponds to the capability of the test

compound to induce beta-cell differentiation.

In a further embodiment (which may be combined with the high-throughput screening as described above) a medium throughput validation is carried out. Therein, the test compound is added to stem cells being cultivated and the DG119-1 and/or DG119-2 expression and/or the insulin production is determined. Following an initial high throughput assay, such as the cell based assay outlined above where e.g. a Pax4 promoter is used as marker for beta-cell regeneration, the activity of candidate molecules to induce beta-cell differentiation is tested in a validation assay comprising adding said compounds to the culture media of the embryoid bodies. Differentiation into insulin-producing cells is then evaluated, e.g. by comparison to wild type and/or Pax4 expressing cells to assess the effectiveness of a compound.

EXAMPLES

A better understanding of the present invention and of its many advantages will be had from the following examples, given by way of illustration.

Examples

Example 1. Identification of DG119 homologues

DG119 homologous proteins and nucleic acid molecules coding therefore are obtainable from vertebrate species, e.g. mammals or fish. Particularly preferred are nucleic acid molecules and proteins encoded thereby comprising human DG119-1 and DG119-2. Human DG119-1 protein was published with GenBank Accession number XP_034000; the corresponding nucleic acid was published with GenBank Accession number XM_034000. Human DG119-2 protein was published with GenBank Accession number NP_872293; the corresponding nucleic acid was published with GenBank Accession number NM_182487. Mouse DG119-1 protein was published with GenBank Accession number NP_796042; the corresponding nucleic acid was published with GenBank Accession number NM_177068. Mouse DG119-2 protein was published with GenBank Accession number NP_766442; the corresponding

nucleic acid was published with GenBank Accession number NM_172854.

To identify possible zebrafish ortologues of human and mouse DG119-1 and DG119-2, sequence databases (NCBI non redundant protein database
5 [\[ftp://ftp.ncbi.nih.gov/blast/db\]](ftp://ftp.ncbi.nih.gov/blast/db), EST section of NCBI genbank (see Boguski
et al., 1993, Nat Genet. 4(4):332-3). dbEST--database for "expressed
sequence tags"], and zebrafish genome draft assembly 2
[\[http://www.ensembl.org/Danio_rerio/ \]](http://www.ensembl.org/Danio_rerio/)) were searched using the blastall
10 programm (version 2.2.6, Altschul et al. 1997, Nucleic Acids Res. 25:3389-
3402). Starting from the blast hits candidate genes were assembled and
translated as necessary using the programm genewise (version 2.2.0, see
<http://www.ebi.ac.uk/Wise2/>), getorf, est2genome and showseq (from the
EMBOSS package version 2.7.1, see
<http://www.hgmp.mrc.ac.uk/Software/EMBOSS/>). The resulting candidate
15 protein sequences were compared to similar mouse and human proteins in
multiple alignments made with the clustalw programm (version 1.83, see
Thompson et al., 1994, Nucleic Acids Research, 22:4673-4680) to verify the
homology to mouse DG119. If available from the assembly data, translation
start sites were selected for antisense oligonucleotides targeting. Otherwise
20 splice donor sites identified by alignment of zebrafish EST data or mouse
protein data to zebrafish genomic sequence were used for antisense
oligonucleotides targeting.

The zebrafish DG119-1a, DG119-1b, DG119-2a, and DG119-2b sequences
25 are shown in FIGURE 6A-D, respectively. An alignment of DG119 from fish,
mouse, and human is shown in FIGURE 7.

Example 2. Loss-of-function experiments in zebrafish

30 To study to effect of DG119 on pancreatic function, several experiments
were performed in zebrafish. Zebrafish were raised, maintained, and crossed
as described (see, Westerfield, 1995, The Zebrafish Book. Eugene, Oregon:
Univ.of Oregon Press). Staging was performed according to Kimmel et al.,

1995, Dev Dyn 203:253-310. Development of zebrafish embryos was carried out at 28 degrees Celsius. The age of embryos is indicated as hours post fertilization (hpf), the age of larvae as days post fertilization (dpf). Zebrafish carrying the transgene with insulin regulatory sequences linked to a fluorescent protein cDNA were used for the experiments. As control, progeny of crosses between AB and TL strain fish (Westerfield et al., supra; <http://www.zfin.org>) were used for injections.

DG119-1 or DG119-2a or control antisense oligonucleotide were injected into fertilized one-cell stage embryos as described (see, for example, Nasevicius & Ekker, 2000, Nat Genet 26:216-220; Urtishak et al., 2003, Dev Dyn. 228(3):405-413). Injected embryos were analysed at different stages of development or processed for quantitative RT-PCR at 48 hours post fertilization (hpf).

Images of zebrafish embryos were taken using a MZFLIII stereomicroscope (Leica) equipped with epifluorescence. In some cases, two images were taken at the same focal plane in transmitted light and using an EGFP filter, and then superimposed and processed using the Adobe Photoshop program.

Figure 1 is showing the internal organs of 17 d old fish embryos carrying a fluorescent protein. At this stage of development, the fish pancreas includes one bigger islet and several smaller islets, as marked by arrowheads in the figure. Figure 2 clearly shows that the injection of DG119-1 antisense oligonucleotides increases the main islet size in these fish, and FIGURE 3 shows the increased size of the small islets after DG119-1 antisense oligonucleotide injection.

Figure 5A shows the effect of loss-of-function of DG119-2 a on the size of pancreatic islets. The islet size is significantly increased injection of antisense oligonucleotides.

Example 3. Expression of DG119-1 and DG119-2 in blood vessels of the pancreas

The full-length clone for mouse DG119-1 in the vector pCMV-SPORT6 was
5 obtained from the Deutsches Ressourcenzentrum für Genomforschung
GmbH (Berlin, Germany) and used as template for the synthesis of
Digoxigenin (DIG)-labeled antisense RNA probes.

For obtaining the DG 119-2 probe for in situ hybridization, RNA was
10 prepared from mouse embryonic stem cell by using Trizol, according to the
manufactorer's instructions. cDNA was synthesized and PCR was performed
for 35 cycles. The sequences of the PCR primers are as follows:

m1192 forward (SEQ ID NO: 5): 3'-gtgctgctgctgctggttttg-5'

m1192 reverse(SEQ ID NO: 6): 3'- ctgtgggctgggggtattctgc-5'

15 The amplified PCR product was cloned into pCMV-Sport6 vector (purchased
from Invitrogen) and used as template for synthesis of Digoxigenin (DIG)-
labeled antisense RNA probes.

The in situ hybridization is described in Collombat et al., 2003, Genes Dev.
20 2003 17(20):2591-603. For DG119 RNA in situ hybridization on pancreas
sections, mouse embryos of day 17 were collected and tissue isolated in ice-
cold PBS. The tissue was fixed overnight in 4% paraformaldehyd containing
2mM EGTA, incubated in 30% sucrose and embedded in tissue freezing
medium (Leica). Tissue sections were applied to Super Frost Plus slides
25 (Menzel-Gläser), dried at 42°C for 30-60min and stored at -80°C.

The hybridization of the DG119 antisense RNA to the mouse embryonic
pancreas tissue can be localized using anti-digoxigenin antibodies.
Defrosted sections were hybridized with Digoxigenin (DIG)-labeled
30 antisenseRNA probes in a medium containing 50% formamid, 10% dextran
sulphate, 1mg/ml yeast tRNA, 0,02% BSA, 0,02% Ficoll, and 0,02% PVP at
65°C overnight. The sections were washed successively in 50% formamid,
1x SSC at pH 5.3, 01% Tween-20 at 70°C for 100 min, and 100 mM maleic

acid, 150 mM NaCl, 0,1% Tween-20 (MABT) at pH 7.5 at room temperature for 1 hour. The sections were blocked in PBS containing 2% Blocking Reagent (Roche) and 20% goat serum for 1 hour. Anti-DIG antibody (1:2500) and guinea pig anti-insulin antibody (1:1000) were applied overnight in the same solution at room temperature. Tissues were washed thoroughly in MABT for 2 hours, and NTMT (100 mM NaCl, 100 mM Tris-HCl at pH 9.5, 50 mM MgCl₂, 1% Tween-20) for 20 min, stained in NBT/BCIP (Roche) in PBS and rinsed in NTMT.

For immunofluorescence detection of insulin and for staining the nucleus, the pancreas sections were rinsed in PBS for 10min. Then, anti-guinea pig Cy3 conjugated secondary antibody (1:500) and DAPI stain (= 4,6-diamidino-2-phenylindole; 1:10.000) were applied in PBS containing 2% goat serum for 30min at room temperature. DAPI is a fluorochrome that binds to DNA and is used to stain the nucleus in fluorescence microscopy. The pancreas sections were rinsed in PBS for 20min and covered with coverslips. Signals of the in situ hybridization were detected by immunofluorescence microscopy.

The results are presented in Figure 4. Figure 4 shows that both DG119-1 (Figure 4a) and DG119-2 (Figure 4b) are co-localized in pancreatic blood vessels.

Example 4. Loss-of-function of DG119-2a stimulates insulin expression in zebrafish (Quantitative RT-PCR analysis)

RNA was isolated from 20-30 fish embryos according to standard procedures, and quantitative RT-PCR (Taqman analysis) was performed according to standard procedures with primers specific for zebrafish Insulin1 (GenBank Accession Number NP_571131), zf Pdx1 (GenBank Accession Number NP_571518), zf Pax 4 (Accession Number lcl|ctg9534 Zebrafish shotgun assembly V2) and 18S RNA (Accession Number D751553).

Figure 5B shows the effect of loss-of-function of DG119-2a on the expression levels of zebrafish insulin, pdx, and pax4. The expression of insulin is significantly enhanced (3-fold) upon inhibition of DG119-2a.

5 All publications and patents mentioned in the above specification are herein incorporated by reference. Various modifications and variations of the described method and system of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with specific
10 preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

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Claims

1. Use of a DG119-1 product and/or a DG119-1 agonist or of a DG119-2
5 antagonist for the manufacture of a medicament to stimulate and/or
induce the differentiation or development of insulin producing cells from
progenitor cells.
2. The use of claim 1, wherein the progenitor cells are stem cells.
- 10 3. The use of claim 1, wherein the stem cells are embryonic or somatic
stem cells.
4. The use of any one of claims 1-3, wherein the stem cells are of
15 mammalian origin, preferably of human origin.
5. The use of any one of claims 1-4, wherein the progenitor cells have
been transfected with a pancreatic gene, particularly the Pax4 gene.
- 20 6. Use of a DG119-1 product and/or a DG119-1 agonist or of a DG119-2
antagonist for the manufacture of a medicament to promote the
protection survival and/or regeneration of insulin producing cells.
7. The use of claim 6, wherein the insulin producing cells are beta-cells.
- 25 8. The use of claim 6 or 7, wherein the insulin producing cells are of
mammalian origin, preferably of human origin.
9. The use of any one of claims 6-8, wherein the insulin producing cells
30 have been transfected or transduced with a pancreatic gene,
particularly the Pax4 gene.

10. The use of any one of claims 1-9 for the prevention or treatment of a disease caused by, accompanied by or associated with impaired beta-cell function.

5 11. The use of claim 10 for the treatment of beta-cell degeneration in patients suffering from diabetes type I, LADA, or progressed diabetes type II.

10 12. The use of claim 10 for the prevention of beta-cell degeneration in patients at risk to develop beta-cell degeneration, like for example but not limited to patients suffering from diabetes type I or II, or LADA in early stages.

15 13. The use of any one of claims 1-12, wherein the active ingredient is administered to a patient

(i) as a pharmaceutical composition e.g. enterally, parenterally or topically directly to the pancreas,

(ii) via implantation of active ingredient expressing cells, and/or

20 (iii) via gene therapy.

25 14. The use of claim 13, wherein the active ingredient is administered in combination with another pharmaceutical composition useful to treat beta-cell degeneration, for example but not limited to hormones, growth factors, or immune modulating agents.

30 15. The use of any one of claims 1-14, wherein the DG119-1 product is a protein including purified natural, synthetic or recombinant DG119-1 and variants thereof.

16. The use of claim 15 wherein variants are selected from insertion, substitution, deletion variants and/or chemically modified derivatives, for example but not limited to hybrids of DG119-1 and other proteins.

17. The use of claim 15 or 16, wherein the DG119-1 product is selected from proteins or peptides substantially homologous to the human DG119-1 protein having the amino acid sequence published as GenBank Accession Number XP_034000.
18. The use of any one of claims 1-17, wherein the DG119-1 product is a nucleic acid, e.g. RNA and/or DNA encoding a DG119-1 protein product.
19. The use of any one of claims 1-18, wherein the DG119-2 antagonist is selected from DG119-2 fragments, modified DG119-2 proteins, antibodies, and biologically active nucleic acids.
20. The use of any one of claims 1-19, wherein an effective amount of cells treated in vitro with the active ingredient are transplanted to a patient in need.
21. The use of any one of claims 1-20, comprising modifying DG119-1 and/or -2 expression, wherein cells from a patient in need that have been modified to produce and secreted DG119-1 protein product or agonist or a DG119-2 antagonist in vitro are re-implanted into the patient and/or wherein cells of a patient in need are modified to produce and secrete DG119-1 protein product or agonist or a DG119-2 antagonist in vivo.
22. A method for differentiating or regenerating cells into functional pancreatic cells, the method comprising: (a) cultivating cells capable of being differentiated or regenerated into pancreatic cells in the presence of an effective amount of a DG119-1 product or an agonist thereof or a DG119-2 antagonist in vitro (b) allowing the cells to develop, to differentiate and/or to regenerate at least one pancreatic function; and (c) optionally preparing an effective amount of the differentiated or

regenerated pancreatic cells for transplantation into a patient in need thereof.

5 23. The method of claim 22, wherein the patient in need is a human individual.

24. The method of claim 22 or 23, wherein the patient in need has (a) functionally impaired, (b) reduced numbers and/or (c) functionally impaired and reduced numbers of pancreatic cells.

10 25. The method of any one of claims 22-24, wherein said patient in need is a type I diabetic patient or type II diabetic patient or LADA patient.

15 26. The method of any one of claims 22-25, wherein the pancreatic cells are insulin-producing cells.

27. The method of any one of claims 22-26, wherein the pancreatic cells are beta-cells of the pancreatic islets.

20 28. The method of any one of claims 22-27, wherein the cells in step (a) are selected from embryonic stem cells, adult stem cells, somatic stem cells or progenitor cells, preferably derived from pancreatic tissue.

25 29. The method of any one of claims 22-28, wherein the cells in step (a) are of mammalian origin, preferably human origin.

30 30. The method of any one of claims 22-29, wherein the cells in step (b) have at least one pancreatic function selected from insulin production in response to glucose and expression of glucagon.

31. A method for differentiating or regenerating cells into functional pancreatic cells, the method comprising: preparing an effective amount of a DG119-1 product or an agonist thereof or a DG119-2 antagonist or

of cells capable of expressing a DG119-1 product or an agonist thereof or a DG119-2 antagonist for administration to a patient in need thereof.

- 5 32. The method of claim 31, wherein the active ingredient is a protein product.
33. The method of of claim 31, wherein the active ingredient is a nucleic acid.
- 10 34. The method of claim 31, wherein cells have been modified to produce and secrete a DG119-1 product or an agonist thereof or a DG119-2 antagonist and are prepared for transplantation into a suitable location in the patient.
- 15 35. A cell preparation comprising functional pancreatic cells treated with an active ingredient selected from a DG119-1 product or an agonist thereof or a DG119-2 antagonist obtainable by the method of any one of claims 22-30.
- 20 36. A cell preparation comprising cells expressing an active ingredient selected from a DG119-1 product or an agonist thereof or a DG119-2 antagonist obtainable by the method of any one of claims 31-34.
- 25 37. The preparation of claim 35 or 36, which is a pharmaceutical composition.
38. The preparation of any one of claims 35-37 for the treatment or prevention of pancreatic diseases, particularly diabetes.
- 30 39. The preparation of any one of claims 35-38 for administration by transplantation or for use in a medical device.
40. The preparation of any one of claims 35-39, which contains

pharmaceutically acceptable carriers, diluents, and/or additives.

41. The preparation of any one of claims 35-40, which is a diagnostic composition.

5 42. The preparation of any one of claims 35-41, which is a therapeutic composition.

43. The preparation of any one of claims 35-42 for the manufacture of an agent for the regeneration of pancreatic tissues or cells, particularly
10 pancreatic beta cells.

44. The preparation of any one of claims 35-43 for application in vivo.

45. The preparation of any one of claims 35-43 for application in vitro.

15 46. A method for identifying and/or characterizing compounds capable of modulating the differentiation or regeneration of cells into functional pancreatic, particularly insulin-producing cells comprising:
contacting a compound to be tested with cells under conditions wherein
20 the cells are capable of being differentiated or regenerated into functional pancreatic cells in the presence of DG119-1, a DG119-1 agonist and/or a DG119-2 antagonist and determining the effect of the compound on the differentiation process.

25 47. The method of claim 46 comprising transfecting the cells with a DNA construct containing a reporter gene under regulatory control of a gene involved in beta-cell differentiation, contacting said transfected cells with a compound to be tested and determining the activity of the reporter gene.

30 48. A method for identifying and/or characterizing compounds capable of modulating the differentiation or regeneration of cells into functional pancreatic, particularly insulin-producing cells comprising:

contacting a compound to be tested with cells under conditions wherein the cells are capable of being differentiated or regenerated into functional pancreatic cells and determining the effect of the compound on the expression of DG119-1 and/or DG119-2.

5

49. Use of a preparation of cells expressing an active ingredient selected from a DG119-1 product or an agonist thereof or a DG119-2 antagonist for the treatment and prevention of diabetes.

10

50. The use of claim 49 for inducing the regeneration of pancreatic cells.

51. The use of claim 50, wherein pancreatic cells are beta-cells of the islets.

15

52. Use of a preparation of cells treated with an active ingredient selected from a DG119-1 product or an agonist thereof or a DG119-2 antagonist for the treatment and/or prevention of diabetes.

20

53. The use of claim 52 wherein the cells are differentiated progenitor cells capable of insulin production.

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Abstract

5 The present invention relates generally to methods for preventing and/or
treating pancreatic disorders, particularly those related to diabetes, by
administering a DG119-1 product or an antagonist thereof and/or an
antagonist to a DG119-2 product.

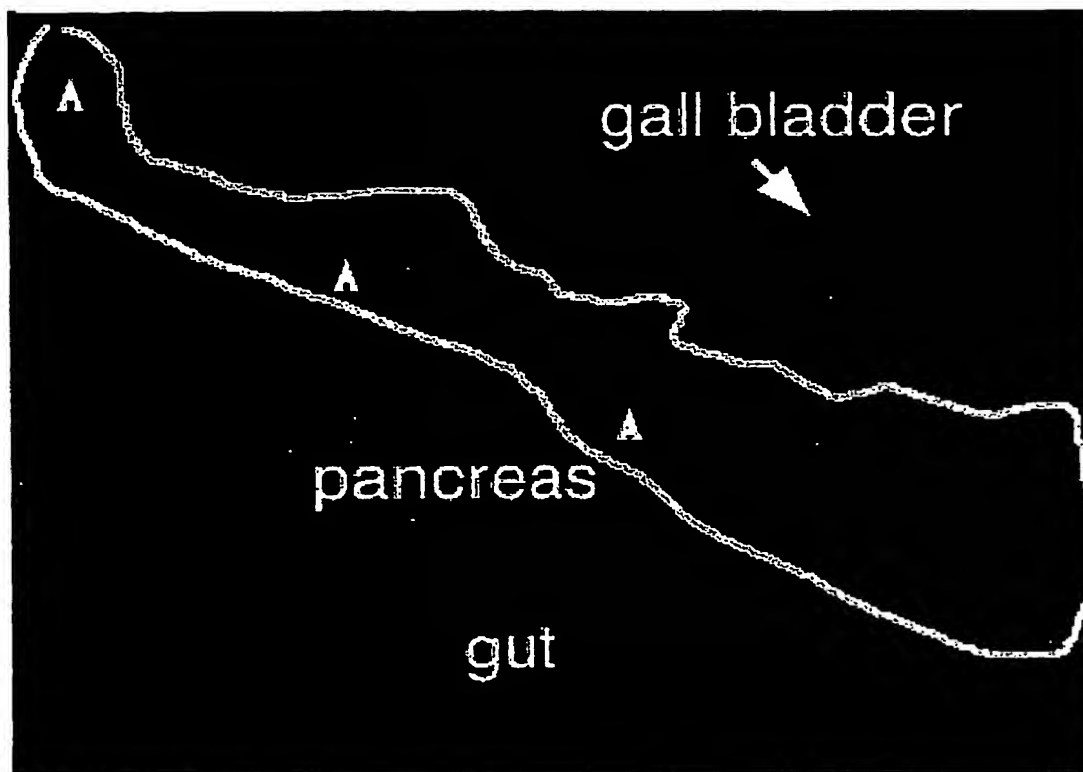
10

Id 28.11.2003

Figure 1. Dev48.
✓

EPO - Munich
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28. Nov. 2003



✓

Control

DG119-1

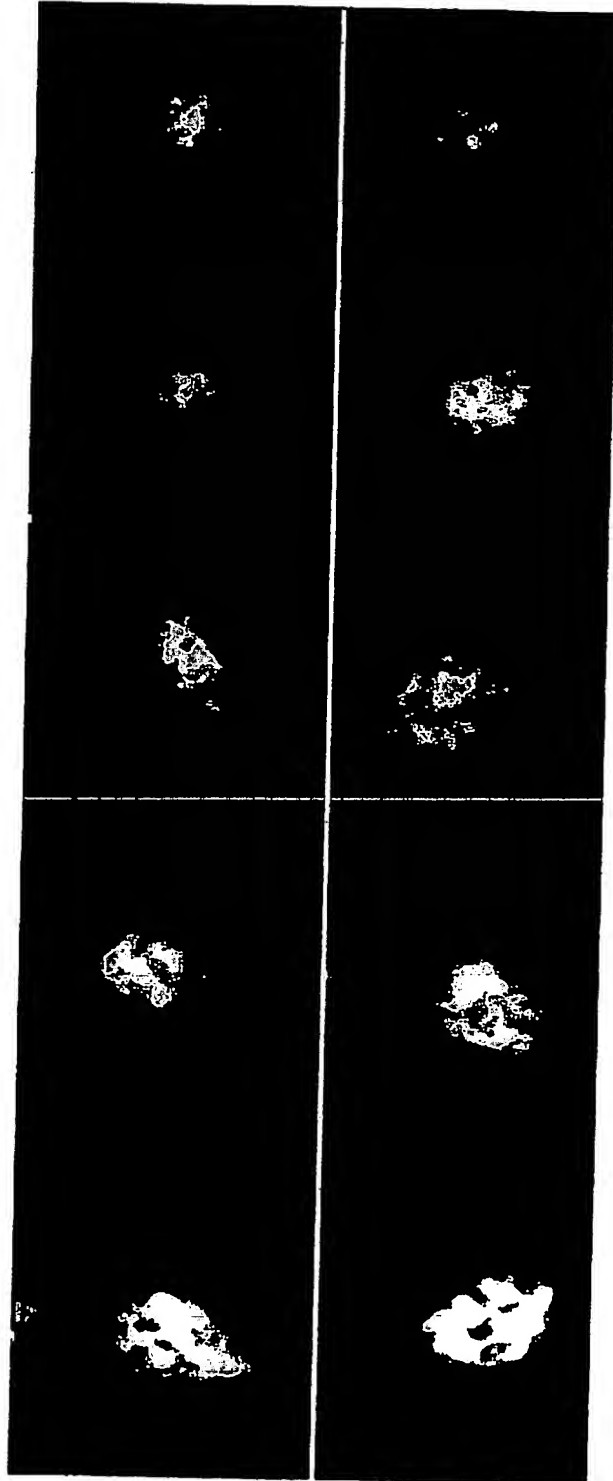


figure 5. dev 10.

✓

Control

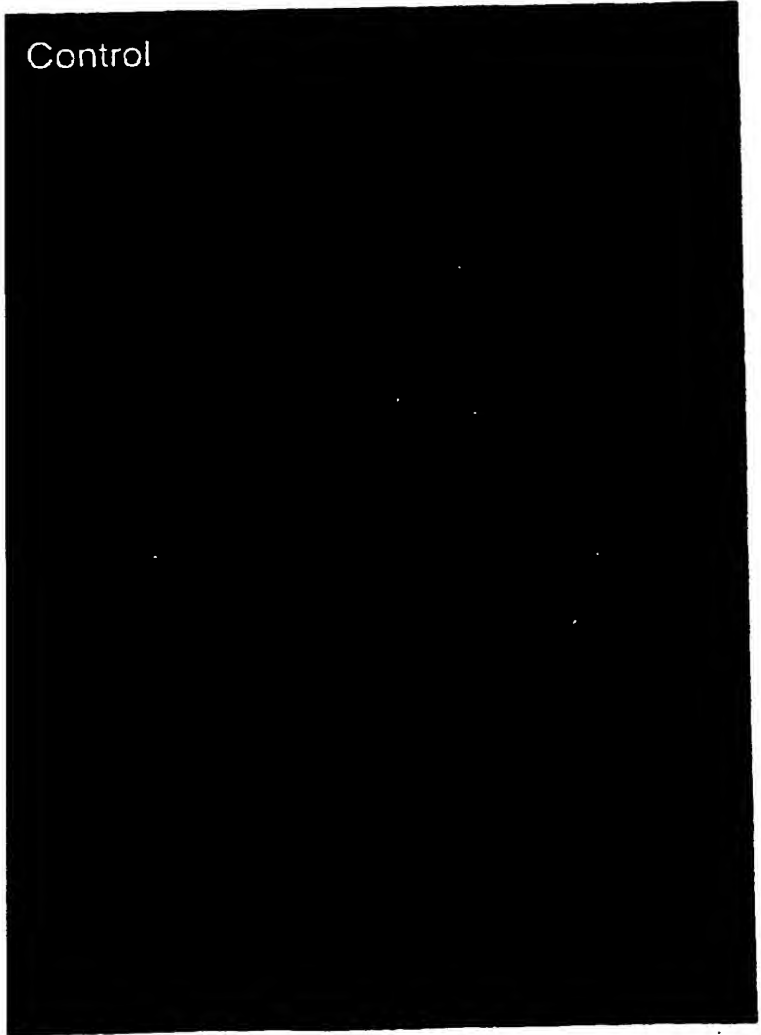


Figure 4a, De

✓

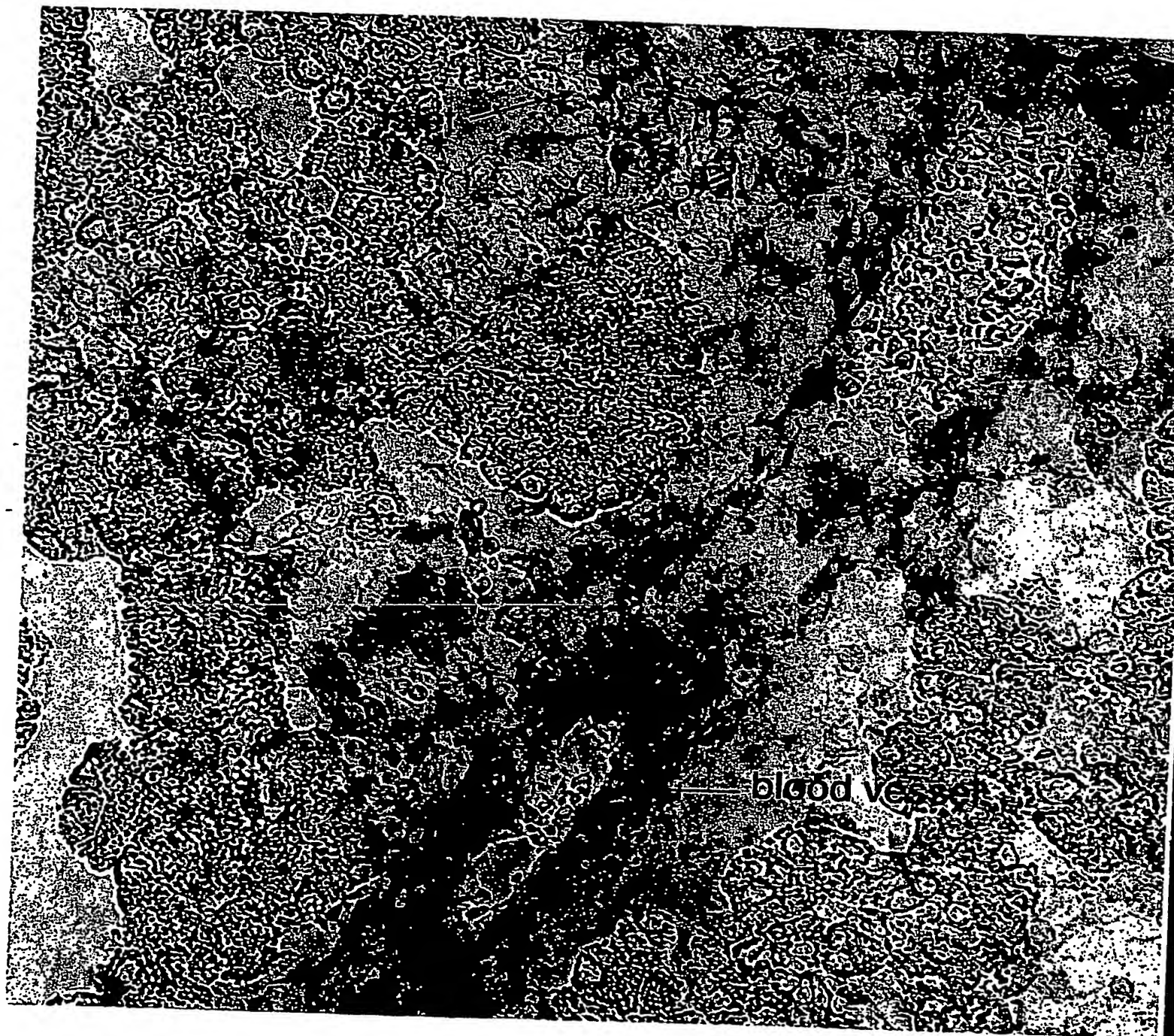


Figure 4b, Dr

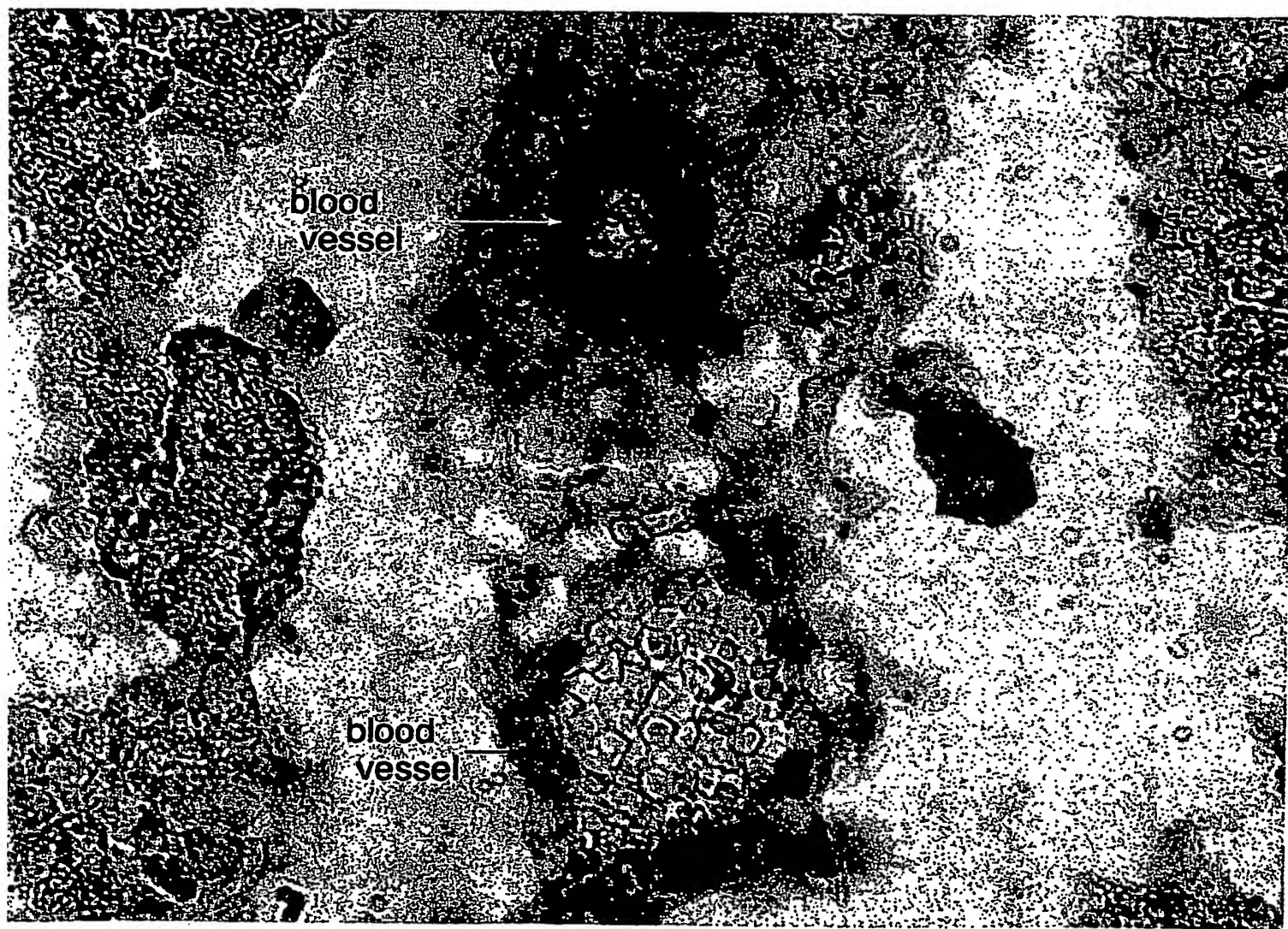
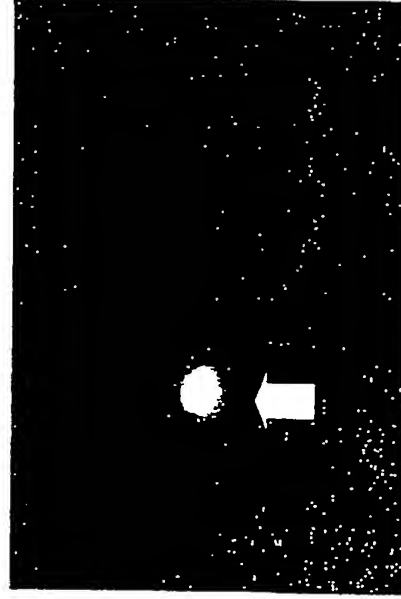


FIGURE 5. DG119-2a



control antisense



DG119-2a antisense

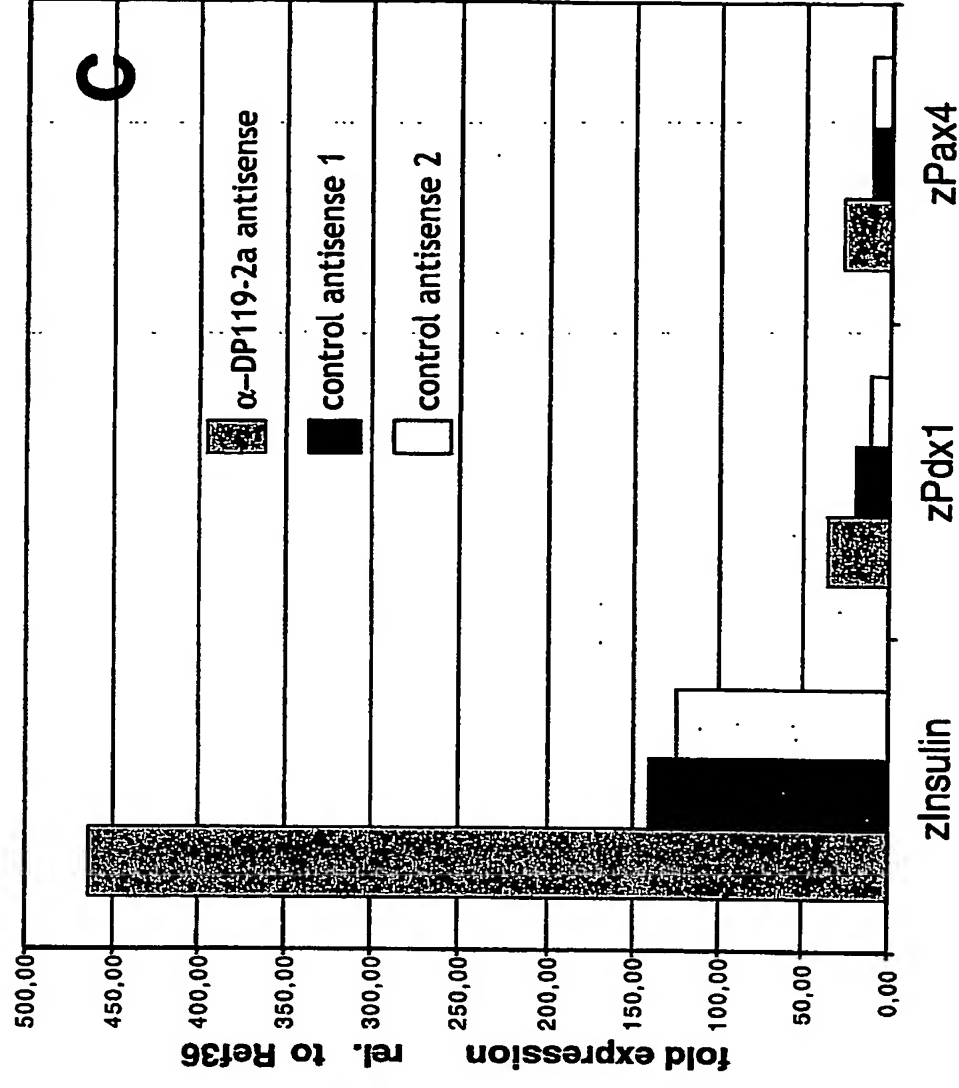


Figure 6. Sequences of zebrafish DG119 proteins.

Figure 6a. Sequence of zebrafish DG119-1a protein (719 amino acids, shown in the one-letter-code)

MTEMKIWCVLLMAFALTSAAPKSHLRLEEKTKDNNDTLQVEIDNQEHILSQLLG DYDKVKALSEGSDCGCKC VVRP
LSASACQRIREGHATPDQDFYTVETITSGPHCKCACIAPPSALNPCEGDFRLKKLRQAGKDNIKLSTILELLEGSFY
GMDLLKLH SVTTKILDRMDTIEKMLNNQTEEKLNITISTSPNPQLSTSSPTTLP SVIQEKSTSLRQONDEAAAFQH
MESKYEEKFVGDILNSGSDLNKATTALQEQEQGRKKQPKITVRGITYYRSDPVD EMDSEKNLKETSA SSVTQTGA
LIKEHLKASTQSTLNTLTPSPTSHSNALTVTESSVGINAHKGEVTTIVMTASVTGSKTDSVTDLTQLSPRVRET LT
TTRTTTKTATT SQPVKRKYSISWDEEEEAVVPEQVEEEKAVKPVVEDKVGEE PQRKPGTAHHQAKTISTVKQ QIRF
SLGMCKDTLATI SEPI THNTYGRNEGAWMKDPLDQDDKIYVTNYYYGNLLEFRNIDVFKQGRFTNSYKLPYNWIG
TGHVVYKGAFYYNRAF SRDIIKFDLRLRYVAAWTMLHDAVFENDDVSSWRWRGNSDMDLAIDESGLWVIYPALDDE
GFLQEVIVLSRLNPTDLSMKRETWTWRTGLRRNRYGNCFIVCGVLYATDSYNQQDTNLSYAFDTHNTQVIPHL PFS
NNYTYVTQIDYNPKERVLYAWDNHGVQVTYNVQFAY

Figure 6b. Sequence of zebrafish DG119-1b protein (594 amino acids, shown in the one-letter-code)

MGLLLYIFCCVFLTRANVEQQATDNTDN RATLEDEMNDQENILTQLIGDYDKVKTLS EGSDCQCKC VVRPMSRSA
CKRIEEAQAKIEDFYTVPEPVTAGPNCKK CACIAPPSALNPCEGDFRKKLQKTGQYDIKLSNIMDLLEERVDNIEK
GKGGQKGARSNQRQEKKKRLSVVCWSLHCRRTQORLLLTLYRCXSVLEPSLOKNAAAFAHTEVQMQQFI PDQR
KYEEKFVGNQGPSKPV LKKSKSEGQEEQHKPAKTKADAKNMSLRSMTFYKANRMEDSEGEERDLI IEDQLHKQGLN
TPVTTPEATVTVTQSTTINLNTQNF TARM SNVTQTQGSVKAMMSSTITTERPTMPTSTTSTSTMTPGTNTTTI
ATPLVVPKQLASVTVGQVSNSYKLPYNWIGTGHVVYSGSFFYNRAF SRDII RFDLRLRYVAAWTTLHDAILEEEEA
PWTWGGHSDIDFSVDESGLWLVYPALDDEGFHQEVIILSKLRASDLQEKSWRTGLRRNRYGNCFVICGVLYAVDS
FERTHANISYAFDTHHTQMI PRLPFINNYTYTTQIDYNPKERMLYAWDNHGVQVTYDVIFAY

Figure 6c. Sequence of zebrafish DG119-2a protein (146 amino acids, shown in the one-letter-code; C-term noncomplete)

MWRIVELVACLLMSSHVSSQSKIFGEEQVRMTSEGS DCRCKCIMRPLTRDACARLTG SVRVEDFYTVETVSSGA
DCKCSCTAPPSSLNPCENEWKREKLKKQAP ELLKLQSMVDLLEGTLP SMDLLKVHSYINKVVSQMNNLEE

Figure 6d. Sequence of zebrafish DG119-2b protein (287 amino acids, shown in the one-letter-code; C-term noncomplete)

MWIYASVLT YLLLLL TRDARSLSKIPGEPEPVKMI SEGSDCRCKCVMRPLSIEACSRLRDGSLRVDDFYTVETVSSG
SDCKCSCTAPPSSLNPCENEWRT EKLXKQAP ELLKLHSMVDLLEGTLYSMDLMKVHAYMNKVVSQMNTLEEVMTIK
TNLTRENEFVRDSVNL SNQLKRYENYS DIMVSIKKEISSLGLQLLQKDAASDSKAQVGTESKKSKEAIKPPNKKP
PAVKPPPKQPK EKPVKPKKEAPAKAAKPAKPDPTTKTKTSVHQTG VIRGITYYKASKSE

Figure 7

DG119 family

		10	20	30	40	50	60
DG119-1	Mm	----- Y	C	Q	WGSTT	TGTSE	D QT E TEDD QNE DNQEN
DG119-1	Hs	----- K R	Y	W SS	TGTSE	D QT	EDET QNE DNQEN
DG119-1a	Dr	----- TE K WC		TS	KS	H R	EETKDNNDT Q E DNQEH
DG119-1b	Dr	----- G	Y CC	C TR N	-----	EQQ	TDNTDNR T EDE DNQEN
DG119-2a	Dr	----- WR	E C	SS	-----	-----	H S
DG119-2b	Dr	----- W Y S	TY	TR	-----	-----	D R
DG119-2	Mm	E	RY Q R	R G	-----	-----	W
DG119-2	Hs	-----	R	SGR	-----	-----	TR
		70	80	90	100	110	120
DG119-1	Mm	SQ	GDYDK K	SEGSDCQCKC	R	GRD CQR	NQG SRKED YT ET TSGSSC
DG119-1	Hs	SQ	GDYDK K	SEGSDCQCKC	R	GRD CQR	N G SRKED YT ET TSGSSC
DG119-1a	Dr	SQ	GDYDK K	SEGSDCQCKC	R	S S CQR	REGH T QD YT ET TSG HC
DG119-1b	Dr	TQ	GDYDK KT	SEGSDCQCKC	R	SRS CKR	EE Q K ED YT E T G NC
DG119-2a	Dr	SQSK	GE-EQ R	TSEGSDCRCKC	R	TRD C R	RTGS R ED YT ET SSG DC
DG119-2b	Dr	S SK	GE E K	SEGSDCRCKC	R	S E CSR	RDGS R DD YT ET SSGSDC
DG119-2	Mm	DSK	SD DQ R	TSEGSDCRCKC	R	SKD CSR	RSGR R ED YT ET SSG DC
DG119-2	Hs	DSK	GD DQ R	TSEGSDCRCKC	R	SKD CSR	RSGR R ED YT ET SSGTDC
----- Cystein rich domain -----							
		130	140	150	160	170	180
DG119-1	Mm	K-C C	S N	CEGD R	QK RE	DSRD	K-----
DG119-1	Hs	K-C C	S N	CEGD R	QK RE	DSQD K G G	GQC GREGT E HKSGK ED
DG119-1a	Dr	K-C C	S N	CEGD R	KK RQ	GKDN	K-----
DG119-1b	Dr	KKC C	S N	CEGD R	KK	QKTGOYD	K-----
DG119-2a	Dr	K-CSCT	SS N	CENEWKREK	KKQ	E K Q	-----
DG119-2b	Dr	K-CSCT	SS N	CENEWRTEK	KKQ	E K H	-----
DG119-2	Mm	R-CSCT	SS N	CENEWK	EK KKQ	E K Q	-----
DG119-2	Hs	R-CSCT	SS N	CENEWK	EK KKQ	E K S	-----
---- Cystein rich domain -----							
		190	200	210	220	230	240
DG119-1	Mm	-----	-----	-----	-----	-----	ST D
DG119-1	Hs	SK	EEG SH	TQT RK	ECTYT	Y YTN	T T3Q SG E ST D
DG119-1a	Dr	-----	-----	-----	-----	-----	ST E
DG119-1b	Dr	-----	-----	-----	-----	-----	SN D
DG119-2a	Dr	-----	-----	-----	-----	-----	S D
DG119-2b	Dr	-----	-----	-----	-----	-----	S D
DG119-2	Mm	-----	-----	-----	-----	-----	S D
DG119-2	Hs	-----	-----	-----	-----	-----	S D
		250	260	270	280	290	300
DG119-1	Mm	EG	YG D	K HS	TTK	G-----	R DK EE
DG119-1	Hs	EG	YG D	K HS	TTK	GR DK EE	EG YG D K HS TTK GR DK EE
DG119-1a	Dr	EGS	YG D	K HS	TTK	D-----	R DT EK
DG119-1b	Dr	EGS	YG D	K HS	TTK	E-----	R DN EK
DG119-2a	Dr	EGT	S D	K HSY	NK	S-----	Q NN EE
DG119-2b	Dr	EGT	YS D	K H Y	NK	S-----	Q NT EE
DG119-2	Mm	EG	YS D	K H Y	QK	S-----	Q NT EE
DG119-2	Hs	-----	-----	-----	-----	-----	-----
		310	320	330	340	350	360
DG119-1	Mm	E--	SKN TKENEQ	KED EE	RTE N-KRGKENCSDNT	ES D RS	QRD
DG119-1	Hs	E--	SKN TKENEQ	KED EE	RTE N-KRGKENCSEN	DS D RS	QRD
DG119-1a	Dr	--	NNQTEEK	NT STS N	-----	TT S	QEKST
DG119-1b	Dr	S--	SGN TKEK	S KGEKGQG	-----	KG RSNQRQEKKKR	S E S QKN
DG119-2a	Dr	-----	-----	-----	-----	-----	-----
DG119-2b	Dr	T	KTN TRENE	RDS N	SNQ KRYENYSD	S KKE SS G Q	QKD
DG119-2	Mm	S--	K N S ENK	KDS HH	SEQ KSYENQS	S KKE SS G Q	QRD
DG119-2	Hs	---	K N SRENE	KDS RH	SEQ RHYENHS	G KKE SR G Q	QKD

		370	380	390	400	410	420
DG119-1	Mm	-----	Y H --EYER	QEET SQQ NS E	RTQ	K QR	QRQ H
DG119-1	Ha	-----	Y H --EYER	QEET SQQ NS E	QTR	E KSQR	QRQ H
DG119-1a	Dr	-----	S RQON-DE	QH ESKYEK	GD NSGSD	NK TT	QE--QEQQGR
DG119-1b	Dr	-----	HTE- Q QQ	DQRKYEEK	GNQG SK	KKSKSEGQ	EQHK
DG119-2a	Dr	-----	SDSK Q	GTEKKSKE	K NKK	K KQ KEK	K KKE K K
DG119-2b	Dr	-----	SS DSK	QDT GGQGRD	NKYGS QKS	SDKG K	KEK K EK R-KES KGR
DG119-2	Mm	T	TG-TGSK	QDT RKGKD	SKYGS QKS	DRG K	KEK Q EK R-KESGKGS
DG119-2	Ha	T	TG-TGSK	QDT RKGKD	SKYGS QKS	DRG K	KEK Q EK R-KESGKGS

		430	440	450	460	470	480
DG119-1	Mm	RGR SK-----	T RG	TYK K	SEEEND	EEQHDE	SGDSG D
DG119-1	Ha	RGR SQ-----	T RG	TYK K	SEEEND	EEQODE	SGDNG D
DG119-1a	Dr	KKQ -----	K T RG	TYRSD	DE DSEKN	KETS SS	TQTG KEH K S
DG119-1b	Dr	KTK D K-----	N S RS	T YK NR	EDSEGEE-----	R D	EDQ HKQ
DG119-2a	Dr	K D TTKTKTS	HQTG	RG	TYK SKSE-----		
DG119-2b	Dr	Q T R R	QQQ	RG	TYK GRQE	RQE RQE	K DST KGTSW EK K
DG119-2	Mm	Q T K R	QQQ	RG	TYK GKQE-----	TE	DN QGTSW EQ K
DG119-2	Ha	Q T K R	QQQ	RG	TYK GKQE-----	TE	DN QGTSW EQ K

		490	500	510	520	530	540
DG119-1	Mm	ED-- TS TRR	TTR--HT	TTD S Q	SSSE	Q S S SS	E Q SDRE
DG119-1	Ha	NG-- TS TRR	TROGHST	TSN N RT	WSS	Q STSD S	NH S G
DG119-1a	Dr	TQST NT T S	TSHSN	T TESS	G N HKGE	TT T S	TGSKTDS TD TQ S
DG119-1b	Dr	G-- NT TT E	T-- T	TQSTT-	N NTQN	TT R SN	TKQTQGS K SS--
DG119-2a	Dr	E K-- E NS	KHDD R Q	SEGGN	T D TTTT	STSSSTTT	TGTSTSTSTSTST
DG119-2b	Dr	EGRSNS	E NS EQDE	E RSSER	D SGTTH	HS HHHST	T H QC
DG119-2	Mm	EGRSNS	E NS EQDE	E RSSER	D SGTTH	HS HHHST	T H QC
DG119-2	Ha	EGRSNS	E NS EQDE	E RSSER	D SGTTH	HS HHHST	T H QC

		550	560	570	580	590	600
DG119-1	Mm	T QTTT	E TG--	STQ S TT	HT Q	GD	E
DG119-1	Ha	--T QTTS	S D TRES	Q S Q	TT HT TQQ	S RE	E HT
DG119-1a	Dr	R RET	TTTTRTTKT	--TTSQ	KRKYS	SWDEEEE	--EQ EEEK K
DG119-1b	Dr	---T TTER	T TS--	TTSTST	T GTNTTT	T	---KQ R---
DG119-2a	Dr	TTT S					
DG119-2b	Dr	HNK	S RRWQTT	SR	GWSNCR	RWR G T	QS T QSR R S G SEWTW
DG119-2	Mm	HNK	S RRWQTT	SR	GWSNCR	RWR G T	QS T QSR R S G SEWTW
DG119-2	Ha	HNK	S RRWQTT	SR	GWSNCR	RWR G T	QS T QSR R S G SEWTW

		610	620	630	640	650	660
DG119-1	Mm	DT GTD	EEEGT	GQE TS G	S EEEDD	RN G	
DG119-1	Ha	TT RTDS	GKD	GWGTT	S T S	EEEDD RN	G
DG119-1a	Dr	EDK GEE	QRK GT	HHQ KT	ST KQQ	K S G	
DG119-1b	Dr						
DG119-2a	Dr						
DG119-2b	Dr						
DG119-2	Mm		TT W TE	H E	SQGRED		
DG119-2	Ha	H N CHHHHRH	H Q	TTS	TE	SG E SSQGRE	

		670	680	690	700	710	720
DG119-1	Mm	---RCKDT	ST TG	TTQNTYGRNEG	W KD	KDDR Y	TNYYYGNT E RN EN
DG119-1	Ha	---RCKDT	ST TG	TTQNTYGRNEG	W KD	KDER Y	TNYYYGNT E RN EN
DG119-1a	Dr	---CKDT	T SE	THNTYGRNEG	W KD	DQDDK Y	TNYYYGNV E RN D
DG119-1b	Dr	---CKDT	S SD	THNKYGNNEG	W KD	KGNGK Y	TDYYGNQ E RD DT
DG119-2a	Dr						
DG119-2b	Dr						
DG119-2	Mm	---SCEGT	R D	KHHSYGRNEG	W KD	DDR Y	TNYYYGNS E RN EN
DG119-2	Ha	---SCEGT	R D	RHHSYGRNEG	W KD	RDDR Y	TNYYYGNS E RN EN

|----- Olfactomedin-like domain -----|

Document made available under the Patent Cooperation Treaty (PCT)

International application number: PCT/EP04/013535

International filing date: 29 November 2004 (29.11.2004)

Document type: Certified copy of priority document

Document details: Country/Office: EP
Number: 03 027 514.3
Filing date: 28 November 2003 (28.11.2003)

Date of receipt at the International Bureau: 01 February 2005 (01.02.2005)

Remark: Priority document submitted or transmitted to the International Bureau in compliance with Rule 17.1(a) or (b)



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